

# A Mini View of Polybrominated Diphenyl Ethers (PBDES) Developmental Neurotoxicity and Mechanism

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## ABSTRACT

Polybrominated Diphenyl Ethers (PBDEs) is a widely used brominated flame retardant extensively incorporated into various consumer and industrial products. Due to its high stability and resistance to degradation, PBDE-209 persists in the environment for prolonged periods and bioaccumulates in the food chain, ultimately reaching the human body. An increasing body of research has demonstrated that PBDEs exerts significant neurotoxic effects, particularly impacting neurodevelopment and neural function. The neurotoxicity of PBDEs is primarily mediated through mechanisms mainly including oxidative stress, mitochondrial dysfunction, disruption of calcium homeostasis, alterations of thyroid hormone homeostasis, interference with neural differentiation, migration and neurotoxicity of PBDEs. This review aims to elucidate the neurotoxic mechanisms of PBDEs and explore effective pharmacological countermeasures. PBDEs exposure is inextricably linked to neurodevelopmental toxicity, involving diverse mechanisms that intertwine and interact. While certain compounds and associated targets have been identified to mitigate this neurotoxicity, extensive human clinical studies are still required to further explore their safety and efficacy.

Keywords: PBDEs; Developmental eurotoxicity; Mechanism; Pharmacological countermeasures

## INTRODUCTION

Polybrominated Diphenyl Ethers (PBDEs), extensively utilized in electronics, textiles, and furnishings for its brominated flameretardant properties, exhibits significant chemical stability and resistance to degradation. Consequently, PBDEs persists in the environment over prolonged periods, bioaccumulating through food chains and ultimately accumulating in human tissues. Increasing studies highlight the significant neurotoxic effects of PBDEs, particularly on neurodevelopment and neural function [1,2]. Consequently, the investigation of effective pharmacological agents to prevent or mitigate the neurotoxic effects induced by PBDEs exposure is of critical significance. PBDE's environmental persistence and bioaccumulation have rendered it a global public health concern. Its biomagnification through the food chain results in significant human body burdens, particularly evidenced by high concentrations of PBDEs in blood, adipose tissue, and breast milk [3]. This phenomenon has raised extensive concerns regarding its long-term health impacts. In this review, we extend knowledge from previous studies of neurotoxicity and underlying mechanisms of adverse effects on the nervous system by exposure to PBDEs and explore potential therapeutic agents.

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#### Neurotoxic effects of PBDEs on humans

Increasing studies have substantiated that PBDEs exerts potential neurotoxic effects on the nervous system, detrimentally influencing both neurodevelopment and neural function. Study by Herbstman et al., found PBDEs in 94% of cord blood samples from 297 Baltimore newborns, with BDE-47 as the dominant congener [4]. This finding raised concerns regarding PBDEs exposure and its potential toxic effects. The inaugural epidemiological investigation conducted by Roze et al., elucidated that prenatal PBDEs exposure correlated with adverse developmental outcomes, including reduced fine manipulative abilities and increased attentional deficits [5]. Research by Herbstman et al., established that higher concentrations of BDEs 47, 99, and 100 in cord blood were linked to lower scores on mental and physical development evaluations at 12-48, and 72 months, and with notable correlations in psychomotor and mental development indices, as well as IQ scores [6]. Gason, et al., measured PBDEs concentrations in colostrum from 290 Spanish women and assessed their infants' mental and psychomotor development at 12-18 months [7]. Higher PBDEs levels, especially BDE-209, were marginally associated with lower mental development scores [7]. The 2013 study by Eskenazi, et al., within the CHAMACOS project provided compelling evidence supporting that prenatal and childhood exposures to PBDEs correlate with reduced attention, motor skills, and cognitive abilities in children, highlighting the adverse effects of PBDEs on neurological development in youth [8]. A 2014 USA study linked prenatal PBDEs exposure to reduced IQ and higher ADHD in children. Specifically, a tenfold increase in BDE-47 levels resulted in a 4.5-point IQ decrease and a 3.3-point rise in ADHD scores by age 5 [9]. Prenatal PBDEs concentration was inversely associated with reading skills and FSIQ and positively associated with externalizing behavior problems at age 8 years [10]. A 2018 study found that prenatal and childhood PBDEs exposure significantly affects memory function in early adolescence, with a more pronounced impact on females [11]. The research in China linked low-level prenatal exposure to PBDEs with long-term behavioral issues in children, including somatic complaints and withdrawal. Girls experienced more pronounced sleep and internalizing problems, while boys showed greater attention issues [12]. Liang, et al., studied part of the HOME project in Cincinnati, measured serum PBDEs levels in 230 children at multiple ages and assessed their reading skills at ages 5 and 8 [13]. They found that higher PBDEs concentrations, including BDE-153, BDE-47, BDE-99, BDE-100, and total of these four (Sum4PBDEs), were linked to lower reading scores, although these associations lost statistical significance after adjusting for other factors [13]. The research conducted by Margoliset, et al., Vuong, et al., vielded congruent findings: Prenatal PBDEs exposure could be a risk factor for reading difficulties in children prenatal exposure to PBDEs is linked to changes in neural activity in brain regions important for inhibitory control, suggesting that these exposures might contribute to ADHD symptoms by altering essential neural functions for cognitive and behavioral regulation [14-17]. Gaylord et al., analysis of data from 2001 to 2016 estimated that PBDEs exposure led to a loss of about 162 million IQ points and contributed to 738,000 cases of intellectual disability, underscoring

the significant impact of in utero PBDEs exposure on intellectual outcomes [18]. These studies collectively indicate that prenatal and neonatal exposure to PBDEs is linked to various neurodevelopmental deficits. The relationship between PBDEs exposure and cognitive function is complex and varies across different racial and socioeconomic groups. These findings highlight the need for ongoing research into neurotoxic mechanisms and interventions to mitigate PBDEs exposure risks during critical developmental periods.

#### Neurotoxic effects of PBDEs on animals

Numerous rodent studies have shown that exposure to PBDEs leads to behavioral changes and brain alterations. Eriksson et al., found that a single dose of BDE-47 and BDE-99 given to mice affected their spontaneous behavior and memory performance in a dose-dependent manner, with observed declines in habituation ability as the mice aged [19]. Markowski, et al., found that male mice exposed to decaBDE from birth to day 21 showed reduced grip strength, altered motor activities, increased errors in learning tasks, and reduced responses to a drug challenge [20]. Exposed female mice primarily showed changes in activity levels. Co-exposure to Polystyrene Nanoplastics (PS-NPs) and BDE-47 disrupts the expression of key genes such as GAP43, TUBA1A, SHHa, MBP, CHRNa7, and AChE in juvenile zebrafish [21]. It is noteworthy that BDE-209 exhibited greater neurotoxic effects compared to other flame retardants such as Hexabromocyclododecane (HBCD) and Tetrabromobisphenol-A (TBBPA) [22]. Co-exposure to PS-NPs and BDE-47 adversely affects neuronal development and the cholinergic system, leading to heightened neurotoxic effects in juvenile zebrafish [23]. Perinatal exposure to PBDE-47 led to increased hyperactivity and possible elevated anxiety in adult female rats, indicated by more movement and less time spent in the central zone of an open field test [24]. The aforementioned studies emphatically indicate that PBDEs exposure is linked to significant neurobehavioral detriments.

## LITERATURE REVIEW

#### Mechanism of neurotoxicity induced by PBDEs

Oxidative stress: Oxidative stress is a pivotal mechanism in PBDE-induced neurotoxicity. Early in vitro studies have demonstrated that Tetra-and PentaBDE, as well as PBDE-209, induce toxicity in primary neurons and neuronal cell lines primarily through oxidative stress, autophagy disruption, and apoptosis [25,26]. Li et al., the interaction between autophagy and apoptosis in PBDE-47induced neurotoxicity, revealing that disrupted autophagy exacerbates apoptosis, leading to significant neuronal loss and cognitive deficits in rats and wortmannin can reverse the adverse effects of PBDE-47 [27]. PBDE-47 has been shown to cause overload, lipid peroxidation, and mitochondrial iron damage in neuron-like PC12 cells, which are indicative of oxidative stress and ferroptosis which is a novel form of cell death dependent on iron and Reactive Oxygen Species (ROS) accumulation [28]. Additionally, PBDEs quinone metabolites' nucleophilic and redox properties contribute to their neurotoxicity [29]. Research demonstrates that BDE-47

treatment in mice activates caspase-3-dependent PKC $\delta$ , which increases ROS levels, causing hippocampal inflammation and cognitive deficits. Inhibition of this pathway using Z-DEVD-FMK, N-Acetyl-L-Cysteine (NAC), or DJ-1 overexpression *via* adeno-associated viral vectors reduces ROS and improves cognitive functions, these substances hold potential as potential therapeutic drugs [30]. These study highlighting the critical role of oxidative stress in PBDE-induced neurotoxicity (Figure 1).



Figure 1: Diagram of predominant mechanisms of PBDEs neurotoxicity.

PBDEs induce neurotoxicity *via* interference with neural differentiation and migration, neurotransmitter systems, oxidative stress, mitochondrial dysfunction, disruption of calcium and thyroid hormone homeostasis, and modulation of gut microbiota. (Some cartoon components were derived from Biorender (https://www.biorender.com)).

Mitochondrial dysfunction: Multiple researchers have elucidated that PBDEs, especially PBDE-47, cause neurotoxicity by targeting mitochondrial dysfunction. PBDE-47 disrupts mitochondrial biogenesis by inhibiting the miR-128-3p/PGC-1a pathway, reducing mitochondrial DNA, ATP levels, and translation products in rat hippocampus and PC12 cells, leading to significant mitochondrial dysfunction and neuronal damage [31]. Subsequent investigations reveal that neonatal exposure to PBDE-47 leads to learning and memory deficits in adult rats, accompanied by mitochondrial abnormalities and apoptosis in striatum. PBDE-47 blocks PINK1/Parkin-mediated the mitophagy, causing defective mitochondria to accumulate, exacerbates mitochondrial dysfunction and results in neuronal death by via the activation of AMPK/ULK1 pathway [32]. PBDEs also induce neurotoxicity by activating the mitochondrial apoptosis pathway. This is demonstrated by

PBDE-47 disrupting calcium ion homeostasis, leading to mitochondrial dysfunction and apoptosis in neurons [33].

Disrupting calcium homeostasis: PBDEs, particularly tetra- and penta-BDEs, significantly influence neuronal signaling, chiefly by inhibiting neurotransmitter uptake within synaptosomes and vesicles and disrupting Calcium (Ca<sup>2+</sup>) homeostasis [34]. Research indicates that BDE-47 and its metabolite, 6-OH-BDE-47, enhance vesicular catecholamine release via exocytosis in PC12 pheochromocytoma cells and significantly elevate intracellular Ca<sup>2+</sup> concentrations, primarily through calcium release from the endoplasmic reticulum and mitochondria [35]. DE-71 affected the cholinergic system and locomotor activity by disrupting zebrafish larvae calcium homeostasis [33]. DE-71, BDE-47, and BDE-99 have been observed to inhibit calcium uptake in rat neural tissues, and BDE-47 interferes with the calcium ion (Ca2+) homeostasis in human neuroblastoma SH-SY5Y cells and hippocampal neurons of rats inducing apoptosis [36,37] suggesting that PBDEs may indirectly affect cellular functions by altering Ca<sup>2+</sup> regulatory mechanisms.

Alterations of thyroid hormone homeostasis: Thyroid hormones are essential for brain development, hypothyroidism linked to numerous neuroanatomical and behavioral abnormalities. Numerous studies have elucidated that PBDEs can disrupt thyroid homeostasis, which is linked to neurotoxicity [1,2]. Epidemiological studies have shown exposure to PBDEs associated with lower TSH during pregnancy, positive associations were found between prenatal PBDEs concentrations and children's neurobehavioral issues, including somatic complaints, withdrawal, sleep problems, and internalizing problems in girls, and somatic complaints and attention problems in boys, inverse associations were observed between the sum of BDE-47, -28, -99, -100, and -153 with thyroid hormones [12]. In a study using female Sprague-Dawley rats administered low doses of PBDE-47 orally disrupted thyroid follicle structure and enhanced thyroid tissue apoptosis. PBDE-47 induced Endoplasmic Reticulum (ER) stress and unfolded protein response, leading to autophagy defects and increased apoptosis, indicating that excessive ER stress and autophagy impairment are involved in maternal thyroid injury following perigestational PBDE-47 exposure [38]. Exposure of fetal human Neural Progenitor Cells (hNPCs) to BDE-47 and BDE-99 did not affect proliferation but reduced migration distance and differentiation into neurons and oligodendrocytes, simultaneous exposure with the Thyroid Hormone Receptor (THR) agonist triiodothyronine rescues these effects on migration and differentiation [39]. Human type II Iodothyronine Deiodinase (Dio2), which is crucial for regulating local cerebral Thyroid Hormone (TH) equilibrium by neuroglial cells, is significantly impacted by BDE209. The BDE209-induced degradation of Dio2 and the subsequent loss of its enzymatic activity in neuroglial cells cause TH disequilibrium and further neurotoxicity [40]. Neonatal rat hippocampal neuron-glia co-cultures exposed to BDE-47 or BDE-49 and treated with triiodothyronine (T3: 3-30 nM), NAC: (100  $\mu$ M), or  $\alpha$ -tocopherol (100  $\mu$ M) showed that T3 and antioxidants effectively counteracted PBDE-induced axonal growth inhibition. T3 also prevented PBDE-induced ROS generation and mitochondrial metabolic disruptions, underscoring

protective role in preserving axonal growth against PBDEs effects [41].

Neural cell differentiation and migration: Neural cells differentiation and migration are critical processes in neurodevelopment. Studies have shown that in vitro exposure to BDE-209 inhibits neurite outgrowth and the differentiation of neural stem cells into neurons in a dose-dependent manner, leading to a higher proportion of neural stem cells differentiating into glial cells [42]. Exposure of human neural progenitor cells to 1 µM BDE-47 or BDE-99 reduces the number of cells differentiating into neuron-like or oligodendrocyte-like cells and decreases their migration capacity [39]. Proteomics studies support these findings, showing that BDE-99 induces changes in the levels of cytoskeletal proteins involved in cellular migration, essential for cell migration [43,44]. Exposure of mice to BDE-209 or its hypobrominated metabolites (BDE-203 and BDE-206) during the critical period of rapid Brain Growth and Development (BGS) disrupts the levels of CaMKII, synaptophysin, BDNF, GAP-43, and tau, which are integral to brain development and function, contributing to processes such as neurite outgrowth and synaptic plasticity. This provides evidence that PBDE-209 can impair key components of normal brain maturation [45,46]. Proteomic analysis in humans with PBDEs exposure exhibited differential expression of 697 proteins, part of which involved cell structure [47]. In our study, we found that PBDE-209 inhibits the differentiation of hippocampal neural stem cells. This effect was reversed by IGF-1, which activated the PI3K/AKT and ERK1/2 signaling pathways, effectively countering the negative impact of PBDE-209 on cellular differentiation [48]. Research indicates that PBDEs, particularly PBDE-47 and PBDE-99, alters the expression of ionotropic glutamate receptors, which are critical for synaptic transmission and the regulation of cell differentiation in the central nervous system [49]. In conclusion, PBDEs impact neurodevelopment by inhibiting neurite outgrowth, reducing neural stem cell differentiation into neurons, impeding cell migration, altering cytoskeletal protein levels.

Effect on neurotransmitter systems: PBDEs disrupt the cholinergic system, affecting neurotransmitters by targeting nicotinic and muscarinic receptors crucial for learning and behavior. Exposure to PBDE-99 causing a decreasing density of cholinergic nicotinic receptors and muscarinic cholinergic receptors in hippocampus [50]. Additionally, research showed that binary mixtures 6-OH-PBDE-47 and PCB-47 could enhance inhibitory GABA (A)-related signals while suppressing excitatory  $\alpha$  (4),  $\beta$  (2) nACh-mediated signals [51]. Following exposure to the PBDE mixture DE-71, changes in the GABAergic and glutamatergic systems in the frontal cortex were observed, revealing alterations in key proteins such as GAD67, vGAT, vGlut, and the GABA(A)  $2\alpha$  receptor subunit [52]. Moreover, DE-71 disrupts the nigrostriatal dopamine system, causing dopaminergic cell death, reduced striatal dopamine, impaired dopamine handling, and significant locomotor deficits in mice, indicating PBDEs as potential risk factors for Parkinson's disease and other neurological disorders [53]. This highlights the impact of early developmental exposure to PBDEs

The mechanisms of brominated Others: diphenvl ether neurotoxicity are intricate, and there are several other perspectives that are also meaningful and warrant attention. A recent study explored the impact of developmental exposure to PBDE-47 on gut microbiota and serum metabolites, linking these changes to effects in adult rats. Gestational and neurobehavioral lactational exposure to PBDE-47 resulted in hyperactivity and anxiety-like behaviors. 16S rRNA sequencing revealed significant changes in gut microbiota composition, with decreased genera Ruminococcaceae and Moraxella, and increased Streptococcaceae, Deferribacteraceae, Escherichia-Shigella, Pseudomonas, and Peptococcus [54]. Metabolomic analysis identified shifts in serum metabolites related to amino acid, carbohydrate, nucleotide, xenobiotic, and lipid metabolism. These disruptions were correlated with neurobehavioral abnormalities, suggesting that gut microbiota dysbiosis and serum metabolite alterations mediate the neurobehavioral impairments induced by early-life PBDE-47 exposure [54] this research provides a novel perspective on understanding the mechanisms of PBDE-induced neurotoxicity, but further research is needed to determine whether adjusting the gut microbiota can alleviate toxicity.

## DISCUSSION

A plethora of cognitive neuroscience, neurobehavioral science, and biological research clearly indicate the neurotoxic potential of PBDEs, PBDEs exerting their neurotoxic effects through mechanisms such as oxidative stress, mitochondrial dysfunction, autophagy, disruption of calcium homeostasis, alteration of thyroid hormone homeostasis, neurotransmitter systems, and interference with neural differentiation and migration, even impact on gut microbiota. Neuronal cells are susceptible to oxidative stress, which results in subsequent damage to DNA, proteins, and membrane lipids. The neurotoxicity of PBDEs is characterized by oxidative stress and related cell death, features similar to those observed in ferroptosis [30]. Caspase-3, a critical effector protease in cell death, and DJ-1, a product of the PARK7 gene, both play pivotal roles in the neurotoxicity induced by stress [55]. Benzyloxycarbonyl-Asp-Glu-Val-Aspoxidative fluoromethyl ketone (Z-DEVD-fmk), a specific caspase-3 inhibitor, and NAC, a thiol-containing antioxidant, have been demonstrated as neuroprotective compounds. Adeno-Associated Viral (AAV) vectors, recognized for their efficacy and safety, have been increasingly utilized in gene therapy for treating neurological disorders. NAC, along with ferroptosis inhibitors such as ferrostatin-1-a lipid reactive oxygen species scavenger-and the iron chelator deferoxamine mesylate, has been shown to mitigate the neurotoxicity induced by PBDE-47. Furthermore, the protein DJ-1 is a potential target for gene therapy using AAV vectors to counteract the neurotoxic effects of PBDE-47 [30]. Exposure to PBDE-47 in human neuroblastoma SH-SY5Y cells triggers increased autophagic activity, characterized by the formation of double-membraned ultrastructure, an increase in MDC-positive cells, and elevated levels of autophagy-related proteins LC3-II, Beclin1, and P62. The use of inhibitors such as

NAC and autophagic inhibitor 3-Methyladenine (3-MA) enhances cell viability, indicating their potential neuroprotective effects by mitigating the autophagy-induced cell death, which is likely mediated by oxidative stress [25]. Autophagy impairment leads to apoptosis and cell death, with wortmannin reversing this effect in PBDE-47 cases [27]. Additionally, melatonin improves PBDE-47-induced neurotoxicity by preventing neuronal apoptosis and loss, restoring mitophagic activity and mitochondrial function via the AMPK/mitophagy axis, 32 therefore, both wortmannin and melatonin act as potential therapeutic compounds in alleviating neurotoxicity. As mentioned before PBDE-47 impairs mitochondrial biogenesis by inhibiting the miR-128-3p/PGC-1 $\alpha$  axis resulting in mitochondrial dysfunction, 31 and further research showed the activation of the PGC-1 $\alpha$ /ERR $\alpha$  axis through the compound ZLN005 restores normal mitochondrial function to mitigate neurotoxicity [56]. In vitro investigations with PC12 cells revealed that PBDE-47 disrupts mitochondrial dynamics, but the chemical promoter M1 or adenovirus-mediated overexpression of Mitofusin 2 (Mfn2) effectively mitigated these effects, improves mitochondrial function, prevents apoptosis, and enhances neuronal survival [24]. NMDA receptor plays an important role in neuronal cell death by causing an excessive influx of calcium, which triggers a series of potentially neurotoxic events. MK801 is known to be a potent and selective noncompetitive antagonist of the NMDA receptor, acting as an open-channel blocker at the NMDA receptor-operated ion channel. The introduction of the NMDA receptor blocker (MK801, 3lM) served to protect against an increase in calcium entry and significantly reduced cell apoptosis induced by DE-71, suggesting the involvement of the NMDA receptor [57]. It has been demonstrating that DE-71 impairs locomotor activity and cholinergic neurotransmission in zebrafish larvae through disruption of calcium homeostasis, this effect can be mitigated by the calcium channel agonist ( $\pm$ )-BAY K8644, as evidenced by significant increases in calcium concentrations, ACh contents, and locomotor activity during light-dark transition stimulation compared to exposure to the same concentrations of DE-71 alone [33], extracellular Ca2<sup>+</sup> chelator (EGTA) showed similar effects [36], further research to explore if other calcium channel regulators produce similar effects is needed. Epidemiological studies and biological research proved that neurotoxicity of PBDEs exposure is linked to thyroid hormone disruption [12,38]. Triiodothyronine (T3) protects axonal growth by inhibiting PBDE-induced ROS [41]. The Insulin-like Growth Factor (IGF) system is crucial for regulating growth, particularly during fetal development, and it plays a central role in maintaining metabolic homeostasis. IGF-1 has demonstrated the capacity to reduce PBDE-induced apoptosis while promoting the proliferation and differentiation of neural stem cells in vitro experiments [48]. Additionally, it inhibits oxidative stress, prevents apoptotic cell death, and enhances neurite outgrowth and cell viability in DRG neurons [58]. Tyrosinase can regulate dopamine synthesis, and studies show that using tyrosinase inhibitors such as Phenylthiourea (PTU) can increase dopamine levels, significantly alleviating the hyperactive behavior of zebrafish embryos induced by PBDEs. This effect suggests that modulating tyrosinase activity holds potential as a therapeutic target for preventing and treating neurotoxicity caused by PBDEs [59]. These therapeutic

approaches are potential for preventing the damaging effects of PBDEs exposure, particularly during important developmental stages.

### CONCLUSION

The neurotoxic mechanisms of PBDEs are complex and intertwined. While pharmacological interventions targeting these key mechanisms have demonstrated potential neuroprotective effects, comprehensive research and clinical trials are essential to confirm their effectiveness and safety in addressing PBDEinduced neurotoxicity.

## CONFLICT OF INTREST

The author declares no conflict of interest.

### REFERENCES

- Sun Y, Xu Y, Wu H, Hou J. A critical review on BDE-209: Source, distribution, influencing factors, toxicity, and degradation. Environ Int. 2023;183(6):108410.
- Dingemans MM, van den Berg M, Westerink RH. Neurotoxicity of brominated flame retardants: (In) direct effects of parent and hydroxylated polybrominated diphenyl ethers on the (developing) nervous system. Environ Health Perspect. 2011;119(7):900-907.
- 3. Lan Y, Gao X, Xu H, Li M. 20 years of polybrominated diphenyl ethers on toxicity assessments. Water Res. 2023;249(3):121007.
- Herbstman JB, Sjödin A, Apelberg BJ, Witter FR, Patterson Jr DG, Halden RU, et al. Determinants of prenatal exposure to Poly Chlorinated Biphenyls (PCBs) and Polybrominated Diphenyl Ethers (PBDEs) in an urban population. Environ Health Perspect. 2007;115(12):1794-17800.
- Roze E, Meijer L, Bakker A, van Braeckel KN, Sauer PJ, Bos AF. Prenatal exposure to organohalogens, including brominated flame retardants, influences motor, cognitive, and behavioral performance at school age. Environ Health Perspect. 2009;117(12):1953-1958.
- Herbstman JB, Sjödin A, Kurzon M, Lederman SA, Jones RS, Rauh V, et al. Prenatal exposure to PBDEs and neurodevelopment. Environ Health Perspect. 2010;118(5):712-719.
- Gascon M, Fort M, Martínez D, Carsin AE, Forns J, Grimalt JO, et al. Poly Brominated Diphenyl Ethers (PBDEs) in breast milk and neuropsychological development in infants. Environ Health Perspect. 2012;120(12):1760-1765.
- Eskenazi B, Chevrier J, Rauch SA, Kogut K, Harley KG, Johnson C, et al. In utero and childhood Poly Brominated Diphenyl Ether (PBDE) exposures and neurodevelopment in the CHAMACOS study. Environ Health Perspect. 2013;121(2):257-262.
- 9. Chen A, Yolton K, Rauch SA, Webster GM, Hornung R, Sjödin A, et al. Prenatal polybrominated diphenyl ether exposures and neurodevelopment in US children through 5 years of age: The HOME study. Environ Health Perspect. 2014;122(8):856-862.
- Zhang H, Yolton K, Webster GM, Sjödin A, Calafat AM, Dietrich KN, et al. Prenatal PBDE and PCB exposures and reading, cognition, and externalizing behavior in children. Environ Health Perspect. 2017;125(4):746-752.
- 11. Cowell WJ, Margolis A, Rauh VA, Sjödin A, Jones R, Wang Y, et al. Associations between prenatal and childhood PBDE exposure and early adolescent visual, verbal and working memory. Environ Int. 2018;118(6):9-16.
- 12. Ji H, Liang H, Wang Z, Miao M, Wang X, Zhang X, et al. Associations of prenatal exposures to low levels of Polybrominated

Diphenyl Ether (PBDE) with thyroid hormones in cord plasma and neurobehavioral development in children at 2 and 4 years. Environment international. 2019;131(6):105010.

- 13. Liang H, Vuong AM, Xie C, Webster GM, Sjödin A, Yuan W, et al. Childhood Polybrominated Diphenyl Ether (PBDE) serum concentration and reading ability at ages 5 and 8 years: The home study. Environ Int. 2019;122(6):330-339
- 14. Margolis AE, Banker S, Pagliaccio D, de Water E, Curtin P, Bonilla A, et al. Functional connectivity of the reading network is associated with prenatal polybrominated diphenyl ether concentrations in a community sample of 5 year-old children: A preliminary study. Environ Int. 2020;134(3):105212.
- 15. Vuong AM, Xie C, Jandarov R, Dietrich KN, Zhang H, Sjödin A, et al. Prenatal exposure to a mixture of Persistent Organic Pollutants (POPs) and child reading skills at school age. Int J Hyg Environ Health. 2020;228(5):113527.
- 16. Sussman TJ, Baker BH, Wakhloo AJ, Gillet V, Abdelouahab N, Whittingstall K, et al. the relationship between persistent organic pollutants and attention deficit hyperactivity disorder phenotypes: Evidence from task-based neural activity in an observational study of a community sample of Canadian mother-child dyads. Environ Res. 2022;206(4):112593.
- Amen NE, Eqani SA, Bilal K, Ali N, Rajeh N, Adelman D, et al. Molecularly tracing of children exposure pathways to environmental organic pollutants and the autism spectrum disorder risk. Environ Pollut. 2022;315(5):120381.
- Gaylord A, Osborne G, Ghassabian A, Malits J, Attina T, Trasande L. Trends in neurodevelopmental disability burden due to early life chemical exposure in the USA from 2001 to 2016: A population-based disease burden and cost analysis. Mol Cell Endocrinol. 2020;502(6):110666.
- Eriksson P, Jakobsson E, Fredriksson A. Brominated flame retardants: A novel class of developmental neurotoxicants in our environment? Environ Health Perspect. 2001;109(9):903-908.
- Markowski VP, Miller-Rhodes P, Cheung R, Goeke C, Pecoraro V, Cohen G, et al. Motor deficits, impaired response inhibition, and blunted response to methylphenidate following neonatal exposure to decabromodiphenyl ether. Neurotoxicol Teratol. 2017;63(3):51-59.
- Kozlova EV, Valdez MC, Denys ME, Bishay AE, Krum JM, Rabbani KM, et al. Persistent autism-relevant behavioral phenotype and social neuropeptide alterations in female mice offspring induced by maternal transfer of PBDE congeners in the commercial mixture DE-71. Arch Toxicol. 2022; 96(1):335-365.
- 22. Wang J, Dai GD. Comparative effects of brominated flame retardants BDE-209, TBBPA, and HBCD on neurotoxicity in mice. Chem Resear Toxicol. 2022;35(9):1512-1518.
- 23. Wang Q, Chen G, Tian L, Kong C, Gao D, Chen Y, et al. Neuroand hepato-toxicity of polystyrene nanoplastics and polybrominated diphenyl ethers on early life stages of zebrafish. Sci Total Environ. 2023;857(6):159567.
- 24. Dong L, Li P, Yang K, Liu L, Gao H, Zhou G, et al. Promotion of mitochondrial fusion protects against developmental PBDE-47 neurotoxicity by restoring mitochondrial homeostasis and suppressing excessive apoptosis. Theranostics. 2020;10(3):1245.
- 25. Zhang C, Li P, Zhang S, Lei R, Li B, Wu X, et al. Oxidative stresselicited autophagosome accumulation contributes to human neuroblastoma SH-SY5Y cell death induced by PBDE-47. Environ Toxicol Pharmacol. 2017;56(3):322-328.
- 26. Sun W, Du L, Tang W, Kuang L, Du P, Chen J, et al. PBDE-209 exposure damages learning and memory ability in rats potentially through increased autophagy and apoptosis in the hippocampus neuron. Environ Toxicol Pharmacol. 2017;50(4):151-158.

- Li P, Ma R, Dong L, Liu L, Zhou G, Tian Z, et al. Autophagy impairment contributes to PBDE-47-induced developmental neurotoxicity and its relationship with apoptosis. Theranostics. 2019;9(15):4375.
- Wang Y, Li X, Qu T, Huang F, Xu J, Gao H, et al. Defective ferritinophagy and imbalanced iron metabolism in PBDE-47-triggered neuronal ferroptosis and salvage by Canolol. Sci Total Environ. 2024;934(3):173118.
- Xu L, Wang Y, Song E, Song Y. Nucleophilic and redox properties of polybrominated diphenyl ether derived-quinone/hydroquinone metabolites are responsible for their neurotoxicity. J Hazard Mater. 2021;420(6):126697.
- 30. Zhuang J, Wang S, Shan Q, Zhang ZF, Li MQ, Zheng GH, et al. Adeno-associated virus vector-mediated expression of DJ-1 attenuates learning and memory deficits in 2, 2<sup>-</sup>, 4, 4<sup>-</sup>. Tetra Bromo Diphenyl Ether (BDE-47)-treated mice. J Hazard Mater. 2018;347(3):390-402.
- Tian Z, Li J, Song L, Xie L, Li D, Xia T, et al. PBDE-47 induces impairment of mitochondrial biogenesis and subsequent neurotoxicity through miR-128-3p/PGC-1α axis. Toxicol Sci. 2023;191(1):123-134.
- 32. Dong L, Sun Q, Qiu H, Yang K, Xiao B, Xia T, et al. Melatonin protects against developmental PBDE-47 neurotoxicity by targeting the AMPK/mitophagy axis. J Pineal Res. 2023;75(1):e12871.
- 33. Wang X, Zhao L, Shi Q, Guo Y, Hua J, Han J, et al. DE-71 affected the cholinergic system and locomotor activity *via* disrupting calcium homeostasis in zebrafish larvae. Aquat Toxicol. 2022;250(5):106237.
- 34. Mariussen E, Fonnum F. The effect of brominated flame retardants on neurotransmitter uptake into rat brain synaptosomes and vesicles. Neurochem Int. 2003;43(4-5):533-542.
- 35. Dingemans MM, de Groot A, van Kleef RG, Bergman A, van Den Berg M, Vijverberg HP, et al. Hydroxylation increases the neurotoxic potential of BDE47 to affect exocytosis and calcium homeostasis in PC12 cells. Environ Health Perspect. 2008;116(5):637-643.
- 36. Zhang S, Chen Y, Wu X, Gao H, Ma R, Jiang C, et al. The pivotal role of Ca<sup>2+</sup> homeostasis in PBDE-47-induced neuronal apoptosis. Mol Neurobiol. 2016;53(4):7078-7088.
- Coburn CG, Currás-Collazo MC, Kodavanti PR. *In vitro* effects of environmentally relevant Poly Brominated Diphenyl Ether (PBDE) congeners on calcium buffering mechanisms in rat brain. Neurochem Res. 2008;33(4):355-364.
- Li P. Perigestational exposure to low doses of PBDE47 induces excessive ER stress, defective autophagy and the resultant apoptosis contributing to maternal thyroid toxicity. Sci Total Environ. 2018; 645(4):363-371.
- Schreiber T, Gassmann K, Götz C, Hübenthal U, Moors M, Krause G, et al. Polybrominated diphenyl ethers induce developmental neurotoxicity in a human *in vitro* model: Evidence for endocrine disruption. Environ Health Perspect. 2010;118(4): 572-578.
- 40. Liu M, Yu Z, Yang F, Zhao Z, Zhou M, Wang C, et al. BDE209promoted Dio2 degradation in H4 glioma cells through the autophagy pathway, resulting in hypothyroidism and leading to neurotoxicity. Toxicology. 2023;494(5):153581.
- Chen H, Carty RK, Bautista AC, Hayakawa KA, Lein PJ. Triiodothyronine or antioxidants block the inhibitory effects of BDE-47 and BDE-49 on axonal growth in rat hippocampal neuron-glia co-cultures. Toxics. 2022;10(2):92.
- Zhang C, Liu X, Chen D. Role of brominated diphenly ether-209 in the differentiation of neural stem cells *in vitro*. Int J Dev Neurosci. 2010;28(6):497-502.
- 43. Alm H, Kultima K, Scholz B, Nilsson A, Andrén PE, Fex-Svenningsen Å, et al. Exposure to brominated flame retardant PBDE-99 affects cytoskeletal protein expression in the neonatal mouse cerebral cortex. Neurotoxicology. 2008;29(4):628-637.

- 44. Alm H, Scholz B, Kultima K, Nilsson A, Andren PE, Savitski MM, et al. *In vitro* neurotoxicity of PBDE-99: Immediate and concentrationdependent effects on protein expression in cerebral cortex cells. J Proteome Res. 2010;9(3):1226-1235.
- **45.** Viberg H. Neonatal ontogeny and neurotoxic effect of Decabrominated Diphenyl Ether (PBDE 209) on levels of synaptophysin and tau. Int J Dev Neurosci. 2009;27(5):423-429.
- 46. Viberg H. Exposure to polybrominated diphenyl ethers 203 and 206 during the neonatal brain growth spurt affects proteins important for normal neurodevelopment in mice. Toxicol Sci. 2009;109(2): 306-311.
- 47. Li M, Huo X, Pan Y, Cai H, Dai Y, Xu X. Proteomic evaluation of human umbilical cord tissue exposed to polybrominated diphenyl ethers in an e-waste recycling area. Environ Int. 2018;111(5):362-371.
- Yang Y, He Q, Zhang Z, Qi C, Ding L, Yuan T, et al. Insulin-like growth factor reduced against decabromodiphenyl ether-209-induced neurodevelopmental toxicity *in vivo* and *in vitro*. Hum Exp Toxicol. 2021;40(12):S475-S486.
- Khani L, Martin L, Pułaski Ł. Cellular and physiological mechanisms of halogenated and organophosphorus flame retardant toxicity. Sci Total Environ. 2023; 897(5):165272.
- 50. Viberg H, Fredriksson A, Eriksson P. Neonatal exposure to the brominated flame-retardant, 2, 2', 4, 4', 5-pentabromodiphenyl ether, decreases cholinergic nicotinic receptors in hippocampus and affects spontaneous behaviour in the adult mouse. Environ Toxicol Pharmacol. 2004;17(2):61-65.
- 51. Hendriks HS, Fernandes ECA, Bergman Å, van den Berg M, Westerink RH. PCB-47, PBDE-47, and 6-OH-PBDE-47 differentially modulate human GABAA and  $\alpha 4\beta 2$  nicotinic acetylcholine receptors. Toxicol Sci. 2010;118(2):635-642.
- 52. Bradner JM, Suragh TA, Caudle WM. Alterations to the circuitry of the frontal cortex following exposure to the polybrominated diphenyl ether mixture, DE-71. Toxicology. 2013;312(4):48-55.

- 53. Dreiem A, Okoniewski RJ, Brosch KO, Miller VM, Seegal RF. Polychlorinated biphenyls and polybrominated diphenyl ethers alter striatal dopamine neurochemistry in synaptosomes from developing rats in an additive manner. Toxicol Sci. 2010;118(1):150-159.
- 54. Qiu H, Gao H, Yu F, Xiao B, Li X, Cai B, et al. Perinatal exposure to low-level PBDE-47 programs gut microbiota, host metabolism and neurobehavior in adult rats: An integrated analysis. Sci Total Environ. 2022;825(4):154150.
- 55. Bonifati V, Rizzu P, van Baren MJ, Schaap O, Breedveld GJ, Krieger E, et al. Mutations in the DJ-1 gene associated with autosomal recessive early-onset parkinsonism. Science. 2003;299(5604):256-259.
- 56. Tian Z, Li J, Tang H, Liu W, Hou H, Wang C, et al. ZLN005 alleviates PBDE47 induced impairment of mitochondrial translation and neurotoxicity through PGC-1α/ERRα axis. J Hazard Mater. 2024;471(5):134331.
- 57. Yu K, He Y, Yeung LW, Lam PK, Wu RS, Zhou B. DE-71-induced apoptosis involving intracellular calcium and the Bax-mitochondriacaspase protease pathway in human neuroblastoma cells *in vitro*. Toxicol Sci. 2008;104(2):341-351.
- Bai X, Chen T, Gao Y, Li H, Li Z, Liu Z. The protective effects of insulin-like growth factor-1 on neurochemical phenotypes of dorsal root ganglion neurons with BDE-209-induced neurotoxicity *in vitro*. Toxicol Ind Health. 2017;33(3):250-264.
- 59. Tanaka Y, Shindo A, Dong W, Nakamura T, Ogura K, Nomiyama K, et al. Tyrosinase inhibition prevents non-coplanar polychlorinated biphenyls and polybrominated diphenyl ethers-induced hyperactivity in developing zebrafish: Interaction between pigmentation and neurobehavior. Neurotoxicol Teratol. 2024;104(2):107373.