

## Autoimmune Mechanism for Leishmaniasis

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## DESCRIPTION

A parasite illness called leishmaniasis is prevalent in several tropical and subtropical regions as well as Southern Europe. It is categorized as a Neglected Tropical Disease (NTD). *Leishmania* parasites that get transmitted by bites of phlebotomine sand flies causes leishmaniasis.

A parasite illness that poses a major threat to human health is leishmaniasis. Leishmaniasis is also the third most prevalent cause of mortality among parasitic infections, behind schistosomiasis and malaria. The possible role of Long Non-Coding RNAs (LncRNAs) causing diseases is yet unknown, though. Investigating the differential expression LncRNAs in leishmaniasis was indeed the goal of this analysis. Highthroughput sequencing was used to gather and analyze the serum of leishmaniasis patients as well as healthy people for controls.

Moreover, qPCR was used to find important LncRNAs that were expressed. The analysis indicated that 1692 different mRNAs and 970 different LncRNAs were evaluated in comparison to control groups. The Encyclopedia Of RNA Interactomes (ENCORI) database and bioinformation analysis were then used to find 520 target genes. The bioinformatics analysis showed that the target genes that were differently expressed were enriched in autophagy, the FoxO signalling pathway, the mTOR signalling pathway, apoptosis, and other processes. Nine important LncRNAs were chosen by qPCR from the differentially expressed LncRNAs (*LINC00622, MAPKAPK5-AS1, LINC02289, XPC-AS1, ZFAS1, and SNHG5* had low expression; MALAT1, NUTM2A-AS1, and LINC00963 used to have high expression).

According to this research, distinct LncRNA expressions may have a role in leishmaniasis and offer a fresh perspective on how to diagnose this zoonotic illness. The primary source of human infection is dogs. For a surveillance programme to be useful, canine visceral leishmaniasis must be diagnosed quickly and accurately. The purpose of this research was to evaluate a quick immunochromatographic strip test for the detection of canine visceral leishmaniasis that was based on functionalized coloured particles and a novel recombinant antigenic protein. An internal ELISA assay utilizing the same antigen was used to assess the outcomes. The immunochromatographic strips test demonstrated strong diagnostic accuracy (98%) and specificity (95%), and both tests yielded consistent findings. Lastly, metaanalysis was utilized to compare the findings of published commercial immunochromatographic strips tests with the specificity and sensitivity of the here devised test.

Leishmaniasis may be diagnosed using a variety of laboratory techniques, both to find the parasite and recognize the species of *Leishmania*. Several of the techniques are only accessible in reference labs. Staff from the CDC in the US can help with leishmaniasis tests. Using a microscope, in specific cultures, and using molecular testing, tissue samples from lesions on the skin or in the blood cells (for visceral leishmaniasis) can be tested for the presence of the parasite. Visceral leishmaniasis cases may benefit from blood tests that search for antibodies (an autoimmune reaction) to the parasite; tests that seek for the infection (or its DNA) directly were performed.

Individual choices should be made about treatment. The CDC team is available for consultation by healthcare professionals who are evaluating different strategies. The kind of leishmaniasis, the Leishmania organism that developed it, the possible severity of the disease, and the patient's underlying health are a few examples of variables to take into account.

The World Health Organization (WHO) reports that in 2018, there were infections in 92 different countries and territories. A billion dogs are at danger of contracting leishmaniasis because they reside in locations where it is common. Each year, around one million instances are recorded. Five categories are used to classify the dog population: susceptible (S), latent (L), infectious (I), infected (Q) and uninfected (R). To analyze and stop the transmission of leishmaniasis, a delayed model was created.

The key characteristics of the model include equilibria, positives, stability, reproduction number, and variable sensitivity. The Routh Hurwitz criteria and Lyapunov theory are used to test the model's reliability both locally and globally. Finally, simulations are provided to support the delayed model's theoretical analysis.

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